Abstract. Hydroxyapatite nanoparticles were isolated from a biowaste, buffalo bone, via the thermal decomposition method. The resulting white powdered material was characterized by Fourier Transformed Infrared (FTIR) spectroscopy, X-ray Diffraction (XRD), Scanning Electron Microscopy (SEM), and Energy Dispersive X-ray (EDX) analysis. The FTIR spectra confirmed that a heat treatment of the bone powder at the temperature at or above 1223 K removed the organic moieties leading to the formation of a pure inorganic biominal. The XRD analyses showed that the obtained material was nanocrystalline HAp (nano-HAp) with an average grain diameter of 25 nm, while their rod-shaped particles with their tightly agglomerated morphology were confirmed by the SEM analysis. Besides Calcium (Ca), Phosphorous (P), and Oxygen (O), trace amounts of Aluminum (Al), Magnesium (Mg), Copper (Cu), Zirconium (Zr) and Carbon (C) were also found by EDX analysis. Antibacterial activity of nano-HAp against six standard isolates was investigated by the agar well diffusion method and found to be more susceptible to Acinetobacter baumannii while other standard strains such as Escherichia coli, Pseudomonas aeruginosa, and Staphylococcus aureus showed lesser susceptibility and no antibacterial activity was noticed against Salmonella typhi and Methicillin resistant Staphylococcus aureus (MRSA) with the analysed concentration of nano-HAp suggesting its potential application in biomedical fields.

Keywords: buffalo bone, hydroxyapatite, nanomaterial, antibacterial activity, bioceramic.
contains a high amount of calcium and phosphorous along with a trace amount of other ions namely Na\(^+\), Zn\(^2+\), Mg\(^2+\), K\(^+\), Si\(^4+\), Ba\(^2+\), F\(^-\), CO\(_2\)\(^-\), etc.\(^{28}\) The extent of such ionic species depends on the age, diet intake, and food habits of the animals.\(^{10,30}\) The presence of these elements in HAp plays a crucial role in the lifecycle of hard tissue metabolism as well as their antimicrobial activities.\(^{10,30}\) Recent studies showed that the HAp particles having a diameter less than 100 nm (nano-HAp) with proper stoichiometry, morphology, and purity have drawn great attention to antimicrobial studies.\(^{31,32}\) The antimicrobial properties of nano-HAp can be enhanced by incorporating other metal ions like Ag, Mg, Ni, Zn, Ti, Na, and Ce or Cu into it.\(^4\)

Recently, cerium-substituted HAp synthesized by the sol-gel method was shown to exhibit good antibacterial properties. The Ce\(^4+\) ions amount in the HAp was found to be an important factor in determining its antibacterial properties similar to silver.\(^{33}\) The incorporation of Ni\(^2+\) ions in the HAp structure exhibited inhibitory property in E. coli and S. aureus.\(^{34}\) It is well known that infectious agents are developing resistance against several antibiotics.\(^{35,36}\) Also antibiotics treatments are becoming costly, possess various side effects, and sometimes are not available in varieties.\(^{36,37}\) Hence, in the pharmaceutical field, the nano-HAp extracted from natural sources could be used as a drug to replace conventional antibiotics as well as a novel nanosized platform for efficient drug carrier and bone tissue engineering.\(^{31,38,39}\) However, only a few such studies have been reported so far.\(^{4,31,33}\) Therefore, also as an extension of our recent work concerning the extraction of HAp nano-bioceramic\(^{22}\), the objective of this work has been the extraction of naturally ion-doped nano-HAp from the waste buffalo bone and its characterization for its structure as well as antibacterial properties.

2. Experimental

2.1. Materials and Sample Preparation

The laboratory-grade chemicals required for this research such as acetone ((CH\(_3\))\(_2\)O), ethanol (C\(_2\)H\(_5\)OH), dimethyl sulfonic acid (DMSO), sodium hydroxide (NaOH), buffer tablets, etc. were purchased from Fischer Scientific (India) and were used without further purification. For antibacterial activity, nutrient agar, nutrient broth, and Mueller-Hinton agar of HiMedia were used as a nutritional substrate. Distilled water was used for preparing the solutions media and for cleansing purposes.

The femur part of adult buffalo bone was collected from a local slaughterhouse in Kathmandu. The bone marrow present inside the femur bone was removed manually by using forceps and needles. Then it was washed thoroughly first with water and boiled in water for 3 h. The clean fat-free bone was obtained by washing it with acetone several times and drying in a normal oven at 393 K for 48 h. Thus, obtained bones were pulverized to get powder using a laboratory grinder.

2.2. Extraction of HAp

To remove collagen, proteins, fats, and other impurities present in the raw bone powder, reflux assisted alkaline hydrolysis of the bone powder was carried out where the raw bone was treated with 4M aqueous alkali (NaOH) in a ratio of 1:40 (w/v) and heated at 523 K for 5 h. For the constant temperature and homogeneous stirring of the solution, a paraffin liquid oil bath and hot plate with a magnetic stirrer were applied. As obtained deproteinated and defatted bone powder was then washed to neutral pH and heated at 923 K for 5 h in a muffle furnace to obtain bone ash. The bone ash was further calcined at 1223 K and then at 1373 K for 6 h to study the effect of calcination temperature.\(^{15,40}\)

2.3. Structural Characterization of Biomaterial

The raw bone powder and bone powder heated at different temperatures were investigated using Fourier Transform Infrared (FTIR) spectroscopy using ITRacer 100 (Shimadzu, Japan) operated in ATR mode. The materials were further analyzed for phase detection and structural characterization by X-ray diffraction (XRD) using monochromatic Cu Ka radiation (D2 Phaser Diffractometer, Bruker, Germany). The average crystallite size was calculated from the XRD data applying Debye-Scherer’s equation.

\[
D = \frac{K\lambda}{\beta\cos\theta}
\]

where \(D\) is the average crystallite size, \(K\) the broadening constant, \(\lambda\) the wavelength of Cu Kα radiation (0.15406 nm), \(\beta\) the full-width at half maximum (FWHM) in radians, \(\theta\) is the diffraction angle in degrees.

Surface morphological analysis of the prepared sample was carried out by using a field emission gun scanning electron microscope (FE-SEM, Hitachi, S-7400). The estimation of calcium and phosphorus (Ca/P) elemental ratio was performed with the help of an energy dispersive X-ray spectrometer (EDX-8000, Shimadzu, Japan).

2.4. Bacterial Culture

Pathogenic reference bacterial strains, namely, gram-negative (Escherichia coli (ATCC 25922), Pseudomonas aeruginosa (ATCC 27853)), gram-positive (Staphylococcus aureus (ATCC 29213)), clinical isolates gram-negative (Acinetobacter baumannii, Salmonella
typhi) and clinical isolates gram-positive (Methicillin resistant Staphylococcus aureus (MRSA)) were collected from the Central Department of Microbiology, Kirtipur, Kathmandu Nepal.

2.5. Antibacterial Activity Assessment

The antibacterial properties of the HAp were analyzed by the diffusion method using sterile Mueller-Hinton agar (MHA) using a standard protocol. Fresh culture of the American Type Culture Collection (ATCC) reference bacterial strains ((quality control strains for antimicrobial susceptibility testing ), namely, gram-negative (Escherichia coli (ATCC 25922), Pseudomonas aeruginosa (ATCC 27853)), gram-positive (Staphylococcus aureus (ATCC 29213)), the identified clinical isolates such as gram-negative (Acinetobacter baumannii, Salmonella typhi) and gram-positive (Methicillin-resistant Staphylococcus aureus (MRSA)) were cultured in nutrient agar at 310 K for 24 h. Fresh cultures of these isolates were sub-cultured in nutrient broth (NB) for 3–4 h at 310 K to obtain the turbidity of 0.5 McFarland standard which is equivalent to 1.5·10^8 bacteria/mL as inoculum. The organisms were lawn cultured with the help of sterile cotton swabs in MHA agar. Then with the help of a sterile 5 mm cork-borer, wells were made in the MHA. The HAp suspensions of various concentrations (5, 10, 20, 30, and 50 mg/mL) were prepared by dissolving the respective powder in dimethyl sulfonylic acid (DMSO).

Wells were filled with 50 µL HAp of different concentrations with the help of a micropipette. DMSO was taken as a negative control. After complete diffusion, the plates were carefully placed in an incubator and maintained at 310 K for 24 h. The next day, the zone of inhibition (ZOI) was noted.

3. Results and Discussion

3.1. Structure and Morphology of HAp

The comparison of the FTIR spectra of raw and alkali-treated bone powder samples calcined at 1223 K and 1373 K is shown in Fig. 1. The possible functional groups and their modes of vibration according to the peaks that appeared in FTIR spectra are summarized in Table 1.

Distinct bands due to the presence of phosphate groups (in the region of 1200–900 and 820–500 cm⁻¹)²⁴, hydroxide group (small bands in the region of 3670–3570 and 640–625 cm⁻¹)²⁴ and carbonate group (small band in the region of 1400–1600 and 870–880 cm⁻¹)²⁴ are observed in all type of samples. The observed results are in agreement with those reported in earlier investigations.³,¹⁵,⁴⁵ However, in the case of raw bone powder, phosphate, hydroxide and carbonate bands are not visible, which could be due to various organic-inorganic cross-linked structures present in the raw bone powder. The observed results are in agreement with previous studies.¹⁵,⁴⁶ Furthermore, the bands around 1646 and 1241 cm⁻¹ appear only in a raw bone powder attributed to the organic compounds present in it.¹³,⁴⁶,⁴⁷

In the case of alkali-treated bone powder, only a few absorption bands appeared whereas when alkali-treated bone powder calcined at 1223 K and 1373 K showed broad absorption bands. The observed bands nearly match with the bands of the HAp spectrum reported in the literature.³,⁴⁸ The peaks located around 3560 and 639 cm⁻¹ can be attributed to the stretching and liberation vibration of (OH) group, respectively.⁴⁹

The IR bands located at 1084, 1034, 963, 599, 561 and 472 cm⁻¹ are due to the vibrations of PO₃⁻ groups. Among these bands, the band positioned at 963 cm⁻¹ is assigned to the symmetric stretching mode ν₁(PO₃⁻)⁻²¹ while the peaks located at 1084 and 1034 cm⁻¹ correspond to the asymmetric stretching mode ν₃(PO₄³⁻)⁻⁵⁰. Likewise, the peaks centred at 599 and 561 cm⁻¹ correspond to asymmetric bending modes ν₃(PO₄³⁻)⁻⁵⁰ and the peak at 472 cm⁻¹ corresponds to the symmetric bending mode of ν₄(PO₄³⁻).⁵¹ In addition weak intensity bands located around 2160 and 2043 cm⁻¹ can be assigned to the overtone and combination of ν₁ and ν₃ mode of PO₄³⁻ group.³⁰,⁵¹

The double bands centred at 1449 and 1417 cm⁻¹ along with the single band centred at 875 cm⁻¹ are due to CO₃²⁻ ions.⁴³,⁴⁷ This is an indication of a minor amount of CO₃²⁻ incorporation in the apatite structure constituting the bone mineral.⁴⁷,⁵² The presence of CO₃²⁻ may play a vital role in enhancing its medicinal properties as carbonate-substituted natural HAp was found to be better for bioresorbable bone substitutes.⁴⁶ However, the intensity of such bands gradually decreases when the high-temperature calcination process is applied³¹ indicating more and more loss of carbon content of bone powder at 1373 K.

![Fig. 1. FTIR spectra of raw, alkali-treated, and calcined (at 1223 K and 1373 K after alkali treatment) bone powder](image-url)

Table 1: Observed FTIR bands with their vibrational assignments.

<table>
<thead>
<tr>
<th>Wavenumber (cm⁻¹)</th>
<th>Vibrational Assignment</th>
</tr>
</thead>
<tbody>
<tr>
<td>3670–3570</td>
<td>Hydroxide groups</td>
</tr>
<tr>
<td>3200–2400</td>
<td>Phosphate groups</td>
</tr>
<tr>
<td>1646</td>
<td>Carboxyl groups</td>
</tr>
<tr>
<td>1084</td>
<td>Symmetric stretching</td>
</tr>
<tr>
<td>1034</td>
<td>Asymmetric stretching</td>
</tr>
<tr>
<td>963</td>
<td>Symmetric bending</td>
</tr>
<tr>
<td>639</td>
<td>Liberation vibration</td>
</tr>
<tr>
<td>472</td>
<td>Symmetric bending</td>
</tr>
</tbody>
</table>

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Table 1. Summary of the functional groups and modes of vibration of various peaks of raw, alkali-treated, and calcined (at 1223 K and 1373 K after alkali treatment) bone samples obtained from FTIR spectra

<table>
<thead>
<tr>
<th>Functional groups and mode of vibration</th>
<th>Raw bone powder, cm(^{-1})</th>
<th>Alkali treated, cm(^{-1})</th>
<th>Calcined at 1223 K, cm(^{-1})</th>
<th>Calcined at 1373 K, cm(^{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td>OH (stretching)</td>
<td>3287</td>
<td>3460</td>
<td>3570</td>
<td>3574</td>
</tr>
<tr>
<td>OH (liberation)</td>
<td>-</td>
<td>-</td>
<td>639</td>
<td>635</td>
</tr>
<tr>
<td>(PO(_4^3-)) (stretching)</td>
<td>1007</td>
<td>1030, 969</td>
<td>1084, 1026, 963</td>
<td>1090, 1030, 969</td>
</tr>
<tr>
<td>(PO(_4^3-)) (bending)</td>
<td>607, 554</td>
<td>608, 558</td>
<td>598, 561, 475</td>
<td>594, 564</td>
</tr>
<tr>
<td>CO(_3^2-) (stretching)</td>
<td>1407</td>
<td>1417</td>
<td>1459, 1408</td>
<td>-</td>
</tr>
<tr>
<td>CO(_3^2-) (bending)</td>
<td>874</td>
<td>872</td>
<td>872</td>
<td>-</td>
</tr>
</tbody>
</table>

There are no absorption bands that correspond to organic impurities (N–H and C–H band)\(^{27}\) for the alkali-treated bone powder calcined at 1223 K and 1373 K suggesting that the extracted HAp would be free from organic impurities.\(^{27}\) Furthermore, an increase in the sharpness of bands, especially that of 639, 599 and 561 cm\(^{-1}\) band, suggests a well-crystallized HAp.\(^{51}\) Hence, as-prepared bone powder proved to be crystalline HAp and found the best to be formed when calcined at 1223 K after alkaline treatment as evidenced by the presence of the expected FTIR peaks in this sample.

The phase structure of the isolated HAp was studied by X-ray diffraction and the observed results for an alkali-treated raw bone sample, as well as a calcined alkali-treated bone powder at 1223 K acquired from 2θ values ranging from 20° to 80° are shown in Fig. 2.

![Fig. 2. XRD pattern of only alkali-treated and the calcined bone powder at 1223 K](image)

The degrees of sharpness of the peaks were found to increase gradually on raising the heat treatment temperature informing the increase in crystallinity of the extracted material as the calcination process helps to release the collagens, fats and other impurities that are bound to the bone powders.\(^{55}\)

The results are in line with the former studies reports.\(^{30,47}\) Furthermore, the broad diffraction peaks observed are closely matched to the stoichiometric HAp characterized pattern (based on ICDD 9-432). The slight deviation of the diffraction peaks from standard peaks could be due to metal ions (Na\(^+\), Mg\(^{2+}\), Al\(^{3+}\), Cu\(^{2+}\), Zr\(^{2+}\), etc.) present naturally in the crystal lattice of HAp.\(^{22,56}\)

Among these, some intense peaks are observed at 20 values of 26.44°, 29.52°, 32.26°, 33.29°, 34.49°, 40.31°, 47.16° and 50.07° which correspond to Miller planes (022), (210), (211), (300), (202), (310), (222), and (213), etc.\(^{29,51}\) Thus, the bone powder calcined at 1223 K after alkali treatment contains pure HAp phase and the obtained result is comparable with the previously reported result.\(^{34}\) In addition, the average grain size of the HAp was found to be 25 nm using the Debye-Scherrer equation and hence it could be considered as naturally ion-doped nano-HAp.

The morphology of the HAp obtained after heat treatment of the alkali-treated bone sample at 1223 K is shown in Fig. 3. As can be seen from the SEM images the as-obtained HAp nanoparticles contained a mixture of differently sized rod-shaped crystalline textures with agglomeration which might have been formed due to their high surface energy and the SEM image is comparable with literature images.\(^{16,57}\) The width of the platelets as observed in Fig. 3. is in the range of 100–300 nm, which is far from that observed by XRD (25 nm) due to agglomeration.

Table 2 and Fig. 4. show the elemental composition and EDX spectrum of isolated HAp from alkali-treated bone powder calcined at 1223 K. The powder was found to consist of Ca, P, O, along with a small amount of Al, Mg, Cu, Zr and C. These trace elements make as-obtained HAp as ion-doped HAp and are essential for antimicrobial activity as well as bone growth.\(^{17,58}\) This analysis showed that the HAp obtained after thermal treatment at 1223 K has mostly inorganic phases. Furthermore, the calcium to the phosphorous elemental ratio of the HAp from EDX study was found to be 1.605, which is smaller (i.e., Ca-deficient) compared to an ideal stoichiometric ratio (1.67).\(^{35,34}\) However, the obtained Ca/P ratio was closed to literature data where the Ca/P ratio was reported to be 1.66 and 1.603 for the HAp recovered from a bovine bone\(^{60}\) and pig bone,\(^{2}\) respectively. Similarly, in another study, the ratio of calcium to phosphorous was reported to...
be 1.86 for the HAp extracted from bovine bone, which is higher than that in the stoichiometric material. These discrepancies in the Ca/P elemental ratio of biogenic HAp may be due to different biogenic sources (bone) of HAp as the trace elemental composition in bone depends on the diet intake and age of the animal itself.28,29

![SEM images](image)

**Fig. 3.** Lower (a) and higher (b) magnified SEM images of the HAp powder obtained after alkali treatment followed by calcination of bone powder at 1223 K

<table>
<thead>
<tr>
<th>Element</th>
<th>Weight %</th>
<th>Atomic %</th>
<th>Element</th>
<th>Weight %</th>
<th>Atomic %</th>
<th>Element</th>
<th>Weight %</th>
<th>Atomic %</th>
</tr>
</thead>
<tbody>
<tr>
<td>C</td>
<td>3.79</td>
<td>7.78</td>
<td>Mg</td>
<td>0.13</td>
<td>0.13</td>
<td>Ca</td>
<td>44.61</td>
<td>27.41</td>
</tr>
<tr>
<td>O</td>
<td>12.5</td>
<td>55.85</td>
<td>Al</td>
<td>0.02</td>
<td>0.02</td>
<td>Cu</td>
<td>0.30</td>
<td>0.12</td>
</tr>
<tr>
<td>Na</td>
<td>0.60</td>
<td>0.64</td>
<td>P</td>
<td>27.78</td>
<td>6.36</td>
<td>Zr</td>
<td>2.27</td>
<td>1.69</td>
</tr>
</tbody>
</table>

**Table 2.** Elemental composition of HAp obtained from bone powder calcined at 1223 K determined by EDX analysis

![EDX spectra](image)

**Fig. 4.** EDX spectra of the HAp obtained by heat treatment of the alkali-treated bone powder at 1223 K

### 3.2. Antimicrobial Properties of Nano-HAp

The antibacterial activity of isolated HAp was investigated by using human pathogenic bacteria, namely *Acinetobacter baumannii*, *Salmonella typhi*, Methicillin-resistant *Staphylococcus aureus* (MRSA), *Escherichia coli*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* by the agar well diffusion method. The observed zones of inhibition (ZOI) after 24 h of incubation are indexed in Table 3.

Among the 6 isolates, the nano-HAp was seen more effective against *Acinetobacter baumannii*, which produced ZOI of diameter 16 mm at 30 mg/mL, 14 mm at 10 mg/mL and 20 mg/mL of nano-HAp, respectively. Other standard strains such as *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* produced ZOI diameters of 11, 10, and 9 mm, respectively, only at higher concentrations of nano-HAp, i.e., 50 mg/mL, and below 30 mg/mL of nano-HAp did not show antibacterial activity (no ZOI observed), as shown in Table 1 and Fig. 5. Furthermore, no ZOI was noticed against *Salmonella typhi* and MRSA with the analysed concentration of nano-HAp. A similar result was reported by Balu et al., the susceptibility to *E. coli* and *S. aureus* was studied and ZOI was found to be 13 and 14.5 mm, respectively, at 50 mg/mL of nano-HAp synthesized from cuttlefish bone. Resmim et al. showed a growth inhibitory effect of *S. aureus* up to 81% and 56% by using nano-HAp from the bovine and porcine bone obtained by calcination at 1273 K, respectively.

Although various properties of HAp including antibacterial properties depend on the size, surface area, morphology, porosity, crystallinity, stoichiometry, and types and concentration of ions present in it,6,10,35 many of the researches reported that the antibacterial activity of nano-HAp is enhanced by doping with other elements like Zn, Cu, Ag, Co, Ce, Se, etc. Hence different ions and its concentration present in the nano-HAp may play a crucial role in inhibiting the bacterial growth along with other properties such as morphology, size, etc.6
Table 3. Zone of inhibition formed by nano-HAp on quality control strains and clinically isolated bacterial strains subjected to interaction with different bacteria as indicated

<table>
<thead>
<tr>
<th>Tested bacteria</th>
<th>Gram reaction</th>
<th>ZOI (mm) of nano-HAp at different concentrations (mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>DMSO (–ve control)</td>
<td>5</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa (ATCC 27853)</td>
<td>–ve</td>
<td>–</td>
</tr>
<tr>
<td>Escherichia coli (ATCC 25922)</td>
<td>–ve</td>
<td>–</td>
</tr>
<tr>
<td>Acinetobacter baumannii</td>
<td>–ve</td>
<td>14</td>
</tr>
<tr>
<td>Salmonella typhi</td>
<td>–ve</td>
<td>–</td>
</tr>
<tr>
<td>Staphylococcus aureus (ATCC 29213)</td>
<td>+ve</td>
<td>–</td>
</tr>
<tr>
<td>MRSA</td>
<td>+ve</td>
<td>–</td>
</tr>
</tbody>
</table>

Note: –ve means gram-negative, +ve means gram-positive and ‘–’ means no zone of inhibition

Fig. 5. Photographs of antibacterial test results of nano-HAp: Pseudomonas aeruginosa (a), Escherichia coli (b), Acinetobacter baumannii (c) and Methicillin-resistant Staphylococcus aureus (MRSA) (d)

Moreover, due to the electrostatic attraction between negatively charged bacterial surface and positively charged HAp, different ions present in the HAp may release in the surrounding environment, which might cause precipitation and coagulation of proteins and enzymes present in the bacterial cell as a result, structural damage occurred within the cell wall and bacterial membranes which ultimately leads to the death of the cell and hence microbes. Such germicidal property of metals ions, especially heavy metal ions, is known as an oligodynamic effect. In addition, the different susceptibility of the material to microbial targets is probably due to differences in the microbial cell wall structures. The results indicate that the nano-HAp extracted from biological sources may be further useful to combat bacterial infections by encapsulation of antibiotics following the principle of targeted drug delivery.

4. Conclusions

Alkaline hydrolysis followed by a calcination process was applied to isolate the nano-HAp from waste buffalo bone. This method is cost-effective and environmentally friendly for the isolation of nanoscale biomaterials. The alkaline hydrolysis and calcination at 1223 K removed the organic impurities from the mineral matrix confirming the formation of carbonate-substituted crystalline HAp as suggested by the IR peaks corresponding to $\text{PO}_4^{3-}$, $\text{OH}^-$ and $\text{CO}_3^{2-}$. XRD study revealed that the isolated biomineral was the nanocrystalline HAp.

The presence of Ca, P and O as major elements and Cu, Zr, Na, Mg, Al, and C as trace elements was confirmed by EDX analysis of the nano-HAp. The Ca/P elemental ratio was found to be 1.605, smaller than the ideal stoichiometric value of HAp, which might have been caused by the ion-doping nature of the HAp derived from biogenic sources.

The nano-HAp biomaterial extracted from the bone was found to possess the rod-shaped agglomerated structure as shown by electron microscopic studies. Among the six standard different isolates investigated for antibacterial activity, nano-HAp was found to be more susceptible to Acinetobacter baumannii while other standard strains such as Escherichia coli, Pseudomonas aeruginosa and Staphylococcus aureus showed lesser susceptibility at the applied concentrations of nano-HAp and no antibacterial activity was noticed against Salmonella typhi and MRSA.
The observed antibacterial properties of the isolated nanocrystalline HAp could be attributed to the ion-doped nature of the HAp. Thus, natural ion-doped bioceramic obtained from biogenic sources has the potential to be applied in different biomedical fields including orthopaedic, dental, and implant applications.

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References


Isolation of Antibacterial Nano-Hydroxyapatite Biomaterial from Waste Buffalo Bone and Its Characterization


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